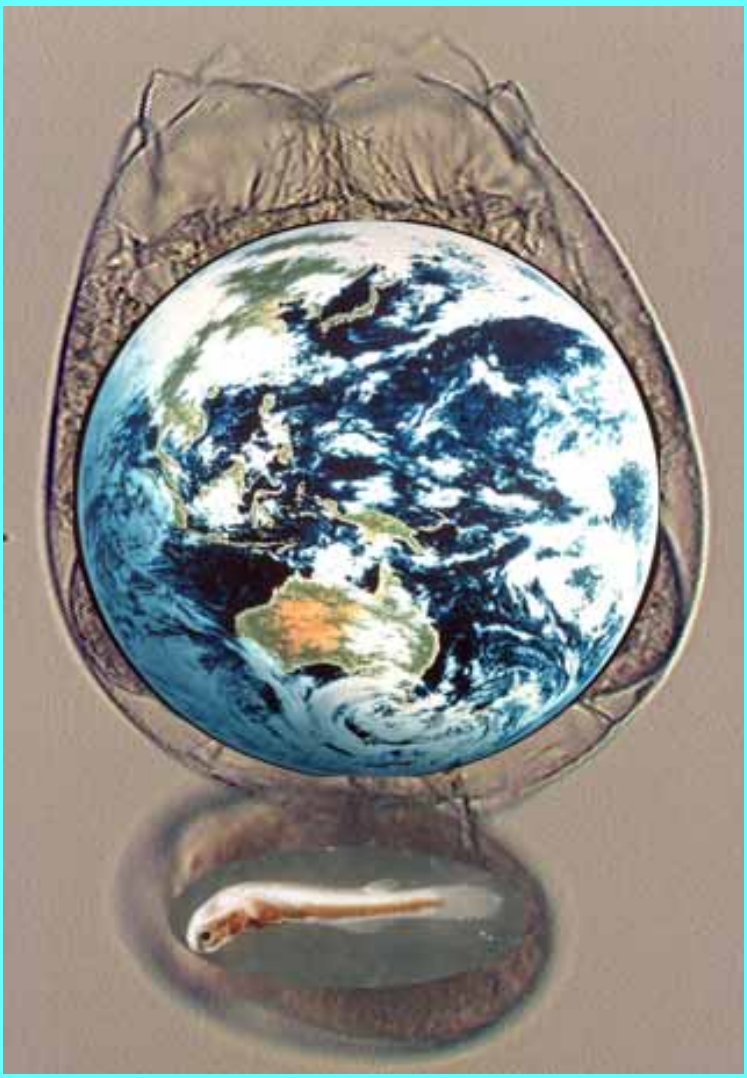
Aquaculture Biology Laboratory

Faculty of Fisheries
Nagasaki University



Professor: Dr. Atsushi Hagiwara (hagiwara@net.nagasaki-u.ac.jp)
Associate Professor: Dr. Yoshitaka Sakakura (sakakura@net.nagasaki-u.ac.jp)
Address: Bunkyo 1-14, Nagasaki 852-8521, Japan
Tel: +81-95-819-2830 or 2823, Fax: +81-95-819-2799

Research projects and plan-do-action of our laboratory

Seedling production (Live feeds & larval rearing) → Aquaculture

The 'live feed'
Rotifer
(0.2 mm BL)

Larva of seven-band grouper
(day 4, 2 mm BL)

Seven-band grouper
(~90 cm BL)

Research Projects

Produce good quality of new target species
→ from empirical to scientific approach

I. Development of live feed planktons

1) Breeding and stock culture of live feed planktons

Culture collection of rotifers …….1

Gene analysis of rotifers …… 2

Resting eggs of rotifers ……..3

Health diagnosis of planktons ……4

Biologically active substances……..5,6

2) New species for live feed …….7

II. Larval rearing

3) Diagnosis of fish health …….8

4) Optimal feeding regime ……9

5) Optimal rearing tank ……..10

III. Publications and Patents …….11,12

Feedback

Novel rearing system

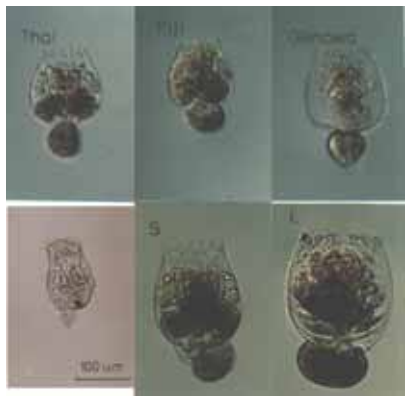
Nagasaki Prefectural Fisheries Experimental Station

Fish farmer, etc…

1. Breeding project of rotifers

Goal: Development of new rotifer strains efficient for larval rearing
Outcomes: ①Cross breeding between rotifers of different strains
②Biological characteristics in cross-breeding strains

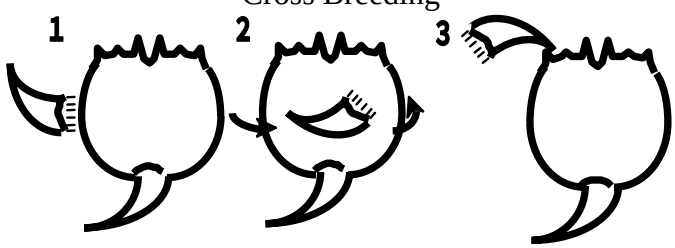
Culture collection of rotifers; they were collected from all around the world



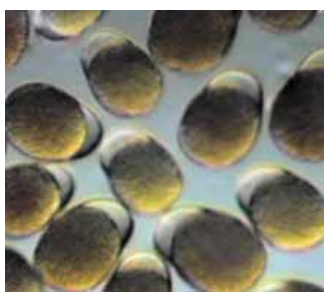
Various strains of rotifers and male (down left)

Strain selection

Cross Breeding

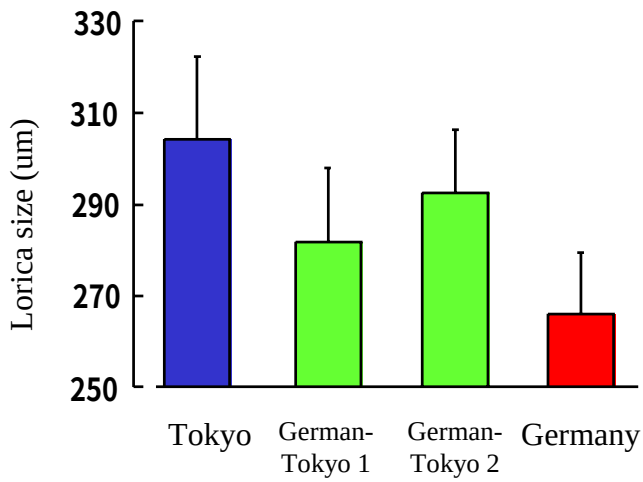


Mating behavior of rotifer



Fertilized eggs
(resting eggs)

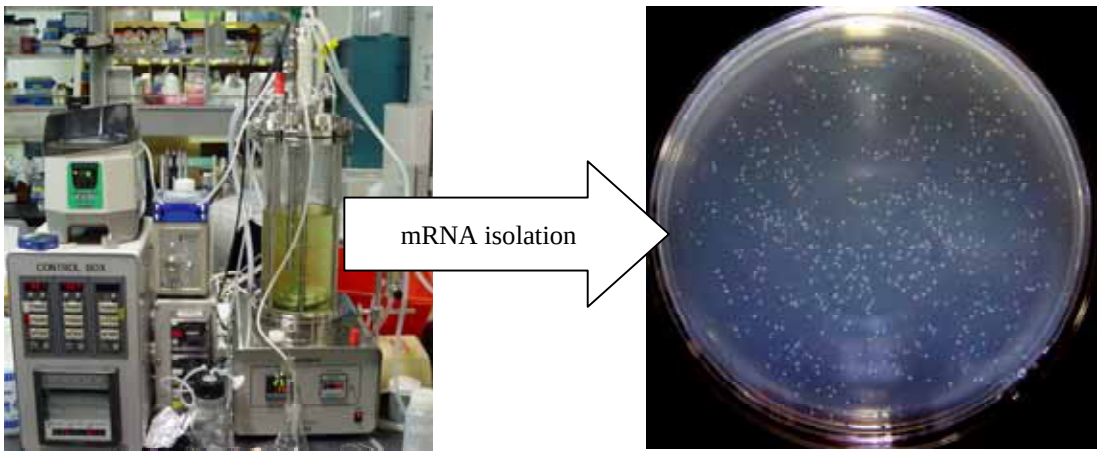
Hatch



2. Construction of expressed sequence tag (EST) and development of transformation method on rotifer

Goal:
1) To construct rotifer EST in order to create cDNA database.
2) To isolate and identify specific genes on rotifer life cycle (male, amictic female and mictic female) by differential display PCR.
3) To create new rotifer strain by chemical manipulation based on the above genetic data.

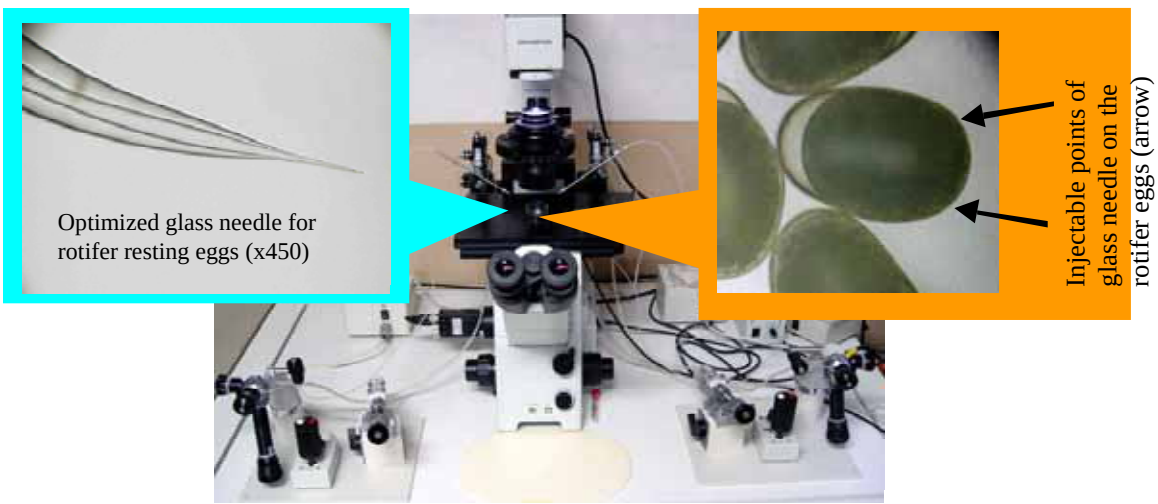
Outcomes:
1) We established an axenic culture method of rotifers by initially using antibiotics. This culture is useful to prevent other organismal DNA in constructing EST (pat. 2003-382155).
2) We isolated mRNA from axenic rotifer culture and constructed cDNA library.
3) We established a transformation technique for rotifer resting eggs using micro manipulator (under patent application and preparation).



Axenic rotifer culture

cDNA library of the rotifer

(plasmid vector with cDNA in *E. coli*)



Micro-manipulator (for chemical injection into rotifer eggs)

3. Rotifer resting eggs: preserved product of live feed for marine fish larvae

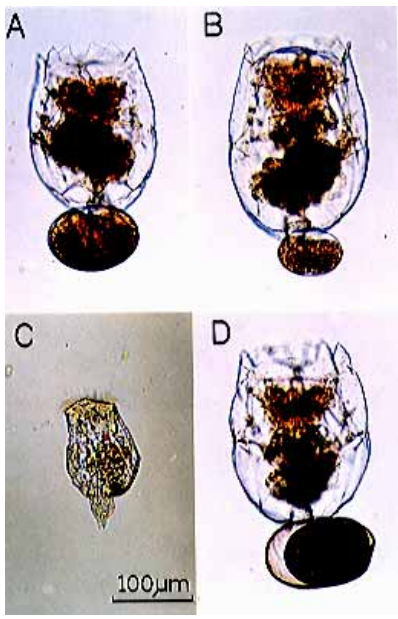
Goal: Development of techniques to mass-produce rotifer resting eggs

Outcomes

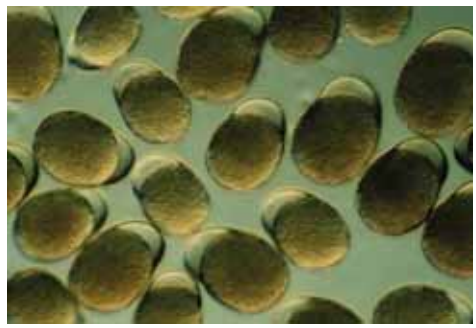
①Rotifer resting egg production by regulating life cycle
(Pat-465050, Pat 2003-323257)

②Artificial sea water for maximizing rotifer resting egg production
(Pat 2003-072203)

③Resting egg production at the order of 10^{10} and preservation by canning



Three female types (A,B,D) and male (C) of marine rotifers



Rotifer resting eggs

Resting egg can containing 10 million resting eggs

Rotifer resting eggs mass-produced in a 50 m^3 tank

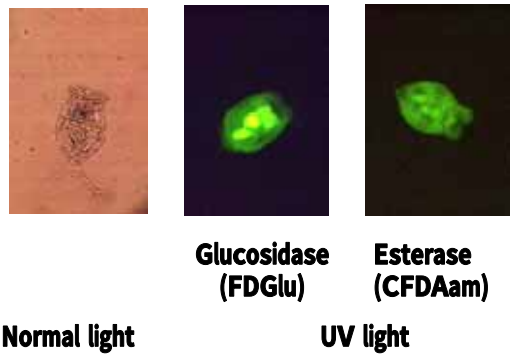
4. Diagnosis of culture status of zooplankton used as live food for rearing fish larvae

Goal : To develop techniques to assess the physiological status of mass cultured marine rotifers used for feeding fish larvae.

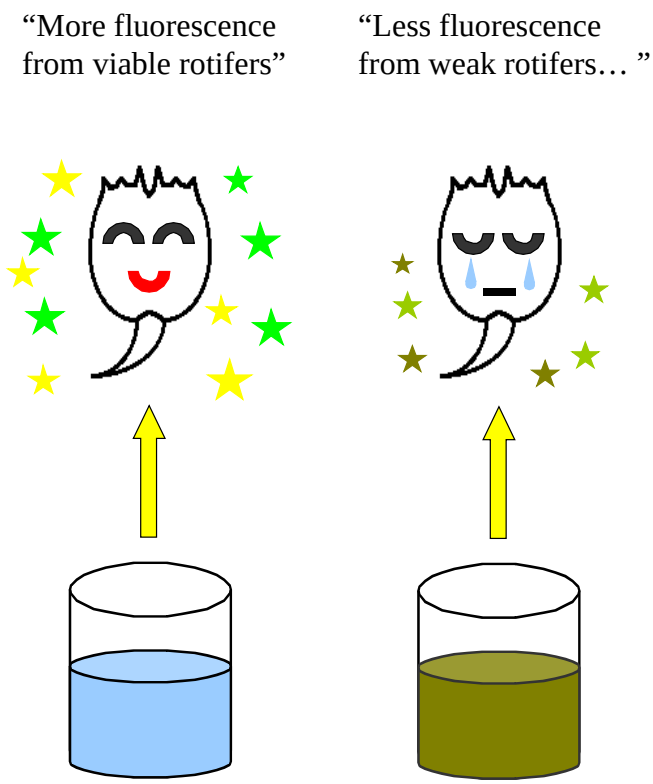
Outcomes:

Environmental changes affect physiological status of rotifers, resulting in the change of life span and fecundity. We confirmed that such changes in demographic parameters correlate with the change in ingestion rate and swimming speed of rotifers.

We further tested the effects of environmental changes on rotifer enzyme activity, and found that it correlates well with rotifer demographic parameters. The use of fluorescent substrate simplifies the process for enzyme activity measurement.



Health check of rotifers



5. Effect of hormone treatments on life history of marine rotifers

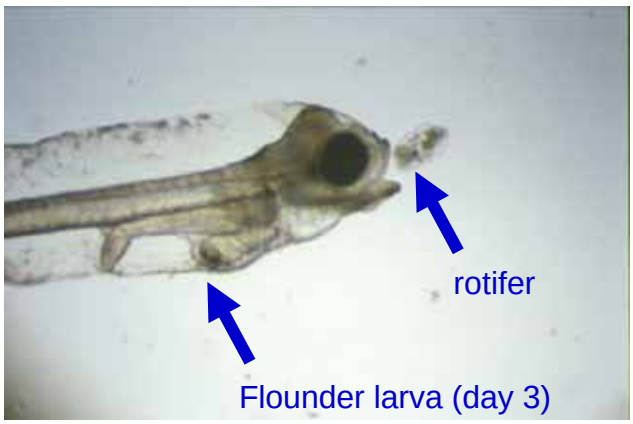
Goal: There are few information about endocrine system of marine rotifers. We examined the effects of the addition of vertebrate and invertebrate hormones on rotifer demographic parameters.

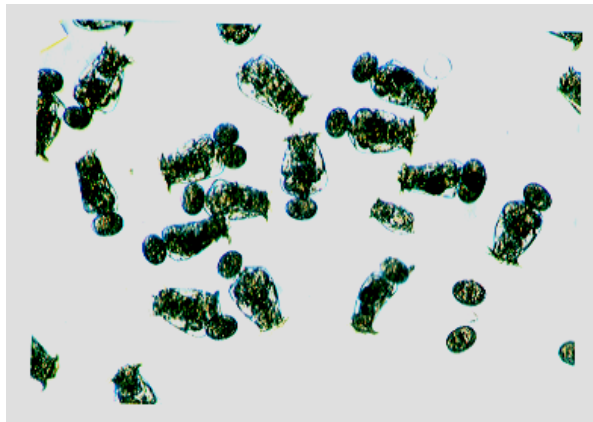
Outcomes

①Among 16 hormones tested (including neurotransmitters), the addition of γ amino-butyric acid (GABA) and porcine growth hormone (GH) promote rotifer population growth. Serotonine (5-HT) and juvenile hormone treatments induced resting egg formation.

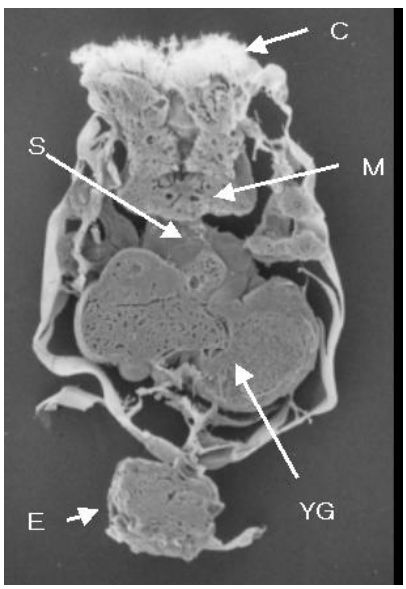
② Effective GABA effects were seen when rotifer cultures are not viable under stressed condition.

③GABA and 5-HT exist in rotifers (HPLC analysis), probably indicating that these function as neurotransmitters in rotifers. GH like substance was found in rotifers, which immuno-react with the antibody of porcine growth hormone.

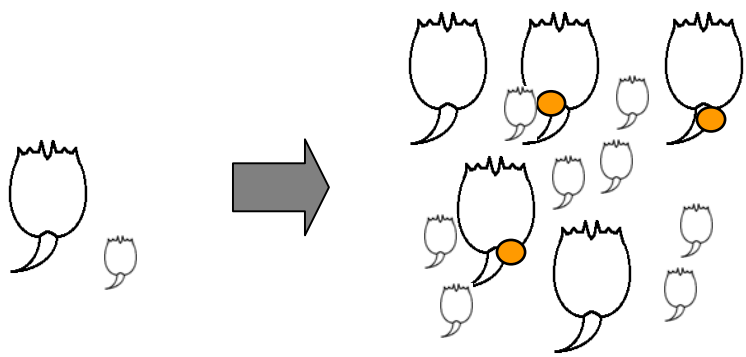




Rotifers in mass cultures



Histological section of marine rotifers; rotifers are metazoeans composed of about 800 cells.



Promotion of rotifer population growth with the addition of GABA and porcine GH

6. Effects of known and suspected endocrine disrupting chemicals (EDCs) on marine zooplankton

Cladoceran
Diaphanosoma celebensis

Copepod
Tigriopus japonicus

Rotifer
Brachionus plicatilis

Goal:

To investigate how EDCs affect zooplankton.

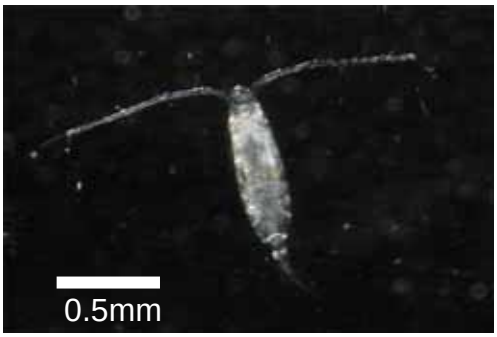
Outcomes:

1) Some EDCs (e.g. 17β-estradiol) increases fecundity of cladoceran.
2) Estrogen and estrogenic compounds affect development of copepods.
3) Some pesticides affect the hatchability of resting eggs of rotifers.

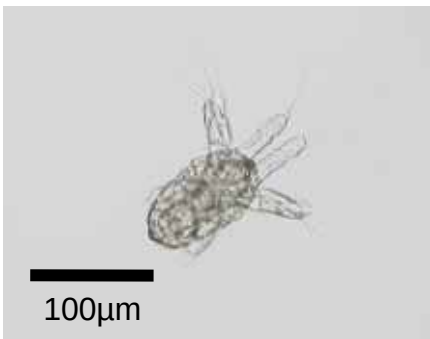
7. Development of culture methods for marine copepods

Goal: Development of novel live feed for fish larva

Outcomes: ①Establishment of culture method for marine copepods
②Long-term culture of copepods from one batch
③Selection of micro algae for marine copepods



Marine copepod, *Acartia tsuensis* (adult)



Nauplius of *Acartia tsuensis*

Small scale (5 L) culture system

20 L-scale copepod culture system with continuous feeding system

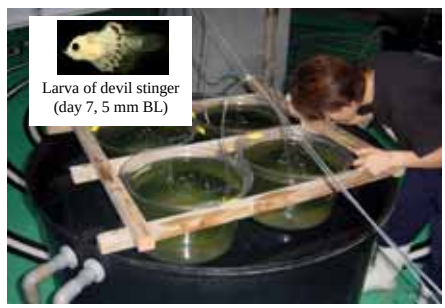
Mass culture system for copepods (400 L)

8. Development of health diagnosis for marine fish larvae

Goal: To establish real-time monitoring system for status of marine fish larvae in the process of seedling production.

Outcomes:
①We found that enzyme activity and behavior of newly hatched larvae have significant positive correlation with health conditions and quality of fish.
②We established individual-base enzyme assay system using clonal lineages of an excellent model fish *Rivulus marmoratus*, that is the only known self-fertilizing vertebrate.
③Real-time diagnosis for health conditions and quality of neonates in the viviparous scorpionfish *Sebastiscus marmoratus* using enzyme activity and behavior (Pat. 3493432).
④We are developing the behavior-analysis computer program, which enables us to observe fish behavior individually, in the collaboration with Industrial Technology Center of Nagasaki Prefecture.

Rearing experiments in the lab.



Rearing experiments in Nagasaki Prefectural Fisheries Experimental Station

Diagnosis of larval quality using enzyme assay and behavioral analysis

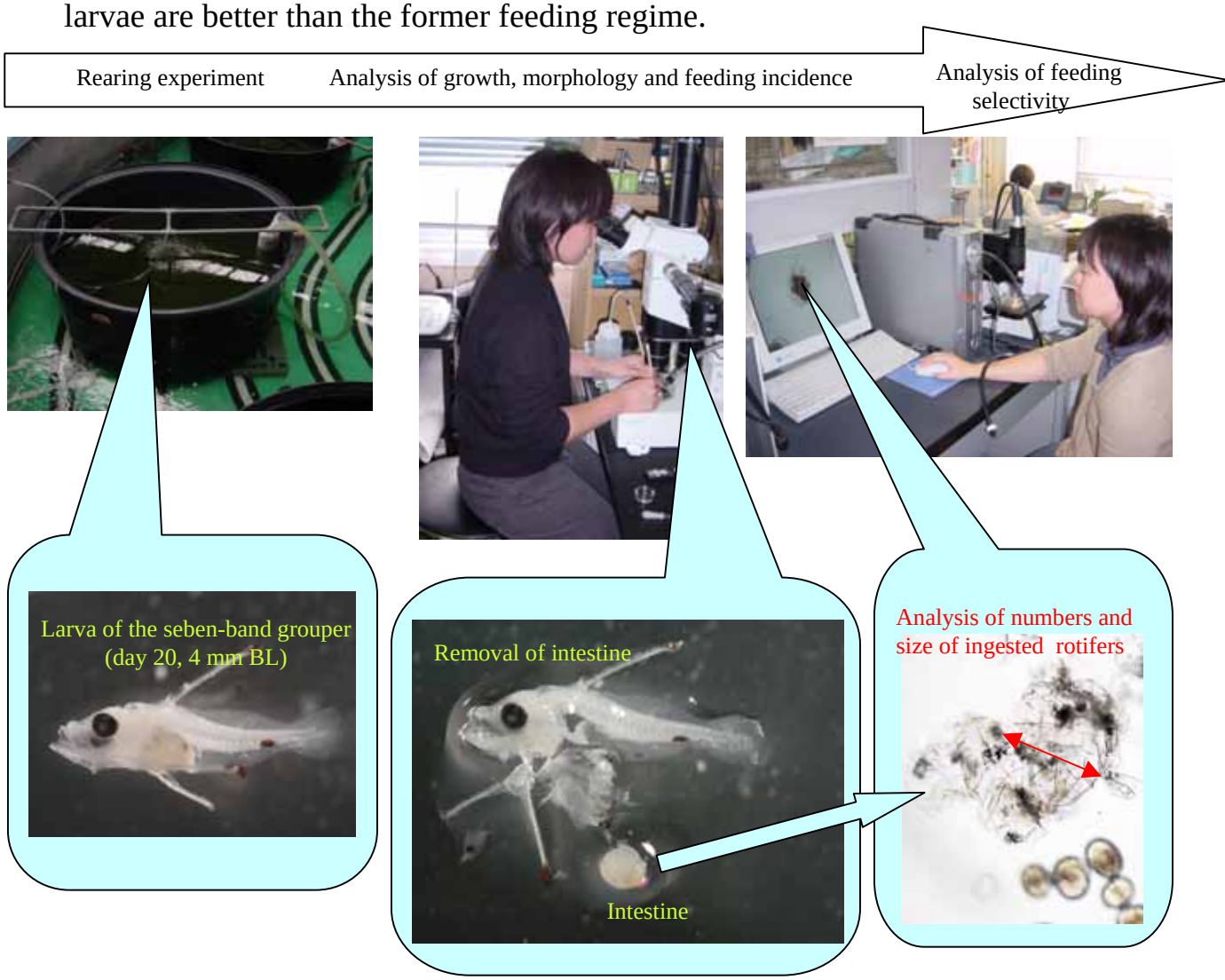
Individual based enzyme assay

Behavioral analysis

9. Development of optimal feeding regime for marine fish larvae

Goal:
To establish the optimal feeding regime during the live feed period of marine fish larvae.

Outcomes:
①We revealed the size-selectivity on live feeds (rotifer and *Artemia*) for the larvae of the seven-band grouper (*Epinephelus septemfasciatus*) and devil stinger (*Inimicus japonicus*).
②We tested a new feeding regime for seven-band grouper based on its feeding selectivity, and found that survival, growth and quality (DHA composition) of larvae are better than the former feeding regime.



Analysis of feeding selectivity for marine fish larvae